

Available Online at www.ijppronline.com International Journal Of Pharma Professional's Research Review Article HPLC: A VERSATILE CHROMATOGRAPHIC APPROACH USED FOR QUALITATIVE AND QUANTITATIVE PURPOSES- A REVIEW



VIKRANT ARYA^{1*}, ANKUR BHARDWAJ², VINIT SHARMA³ ISSN NO:0976-6723 Department of Pharmacognosy, Amar Shaheed Baba Ajit Singh Jujhar Singh Memorial Post Graduate College of Pharmacy, Bela, Ropar, Punjab-140111, India

Abstract

High Performance Liquid Chromatography (HPLC) has been used for the analysis of natural and synthetic compounds. This review gives an overview about some special features involving the mode of operation, mobile phase, detector, column and the flow rate of solvent system employed. This review paper provides useful information regarding the identification, separation and quantification of various natural as well as synthetic compounds.

Keywords: - : HPLC, quantification, phytoconstituents, synthetic

Introduction

A variety of methods are available for analyzing pharmaceutical compounds. High Performance/ Pressure Liquid Chromatography (HPLC) is one of the best methods of choice for analysing a variety of natural and synthetic compounds. It is because it offers high performance over ambient pressure.[1] This method is used for checking the purity of new drug candidates, monitoring changes or in process testing for developing new formulations and quality control or assurance of final drug products. HPLC is a dynamic adsorption process[2] and is a separation technique conducted in the liquid phase in which a sample is separated into its constituent components by distributing between the mobile phase and a stationary phase. HPLC utilises a liquid mobile phase to separate the components of a mixture. The stationary phase can be a liquid or a solid phase. These components are first dissolved in a solvent, and then forced to flow through a chromatographic column under a high pressure.[3]

Correspondence Address:

Vikrant arya

Amar Shaheed Baba Ajit Singh Jujhar Singh Memorial Post Graduate College of Pharmacy, Bela, Ropar, Punjab-140111, India Email: arya.vikrant30@gmail.com Phone:91-09736105832

HPLC acquires a high degree of versatility not found in other chromatographic systems and it has the ability to easily separate a wide variety of chemical mixtures. Some axioms of HPLC are: Sample must be soluble; for separation, analytes must be retained and have differential migration in the column; the mobile phase controls HPLC separation; the final analyte solution should be prepared in the mobile phase.[4]

In this review, we discussed the role of HPLC in the qualitative and quantitative determination of natural phytoconstituents and synthetic drug molecules.

HPLC EVALUATION FOR PHYTOCONSTITUENTS [5-45]

For quality control of herbal products, high performance liquid chromatography (HPLC) is a popular method for the analysis of herbal medicines because it is accurate, precise and not limited by the volatility or stability of the sample compounds. HPLC combined with diode array detector (HPLC/DAD), electrochemical detection (HPLC-ED), mass spectrometer (HPLC /MS) have been successfully employed in qualitative and quantitative determination of various types phytoconstituents including alkaloids, flavonoids, tannins, glycosides, triterpenes, sterols *etc.* A brief description of HPLC conditions for some phytoconstituents were discussed in table 1.

4.

Table	1:	HPLC	conditions	for	some
phytoco	onstit	uents eval	uation[5-45]		

Sr.no	Plant and family name	Chemical constituents evaluated	Method	Mobile phase	Colum n	Detector and wavelength (nm)	Solvent flow rate (mL/min
1	Agrimonia eupatoria, Rosaceae	Procyanidins, kaempferol, 3-O-(6-O- p-coumaroyl)-glucoside and quercetin glycosides	HPLC/DAD/M S	Solvent A- 5% aqueous acetic Solvent B- acetonitrile (2.5%) in aqueous acetic acid in ratio 10:90 Solvent C, acetonitrile	ODS2	Diode array λ_{max} at 280	0.5
2	Fragaria ananassa, Rosaceae	Glycosides of quercetin, kaempferol, cyanidin, pelargonidin and ellagic acid, flavanols, derivatives of p- coumaric acide and ellagitannins	HPLC/DAD/M S	Solvent C- accionatine Solvent A- acetic acid/water Ratio: 2:98 v/v Solvent B- acetic acid/acetonitrile/water Ratio: 2:50:48 v/v/v	C-18	Coulometric array λ _{max} at 260, 360 and 500	0.25
3	Garcinia mangostana, Guttiferae	α-mangostin	RP-HPLC	Solvent A -0.1 %v/v ortho phosphoric acid Solvent B- acetonitrile	BDS C- 18	UV-VIS λ_{max} at 320	1.0
4	Punica granatum, Punicaceae	Phytoestrogenic flavonoids anthocyanins and ellagic acid	HPLC	Solvent A -2.5% v/v, solution of acetic acid in water Solvent B -2.5% v/v solution of acetic acid in methanol	RP C- 18	UV-VIS λ _{max} at 510	1.0
5	Capsicum annuum, Solanaceae	Arbutin	HPLC	Methanol: water Ratio: 90:10	C-18	UV-VIS λ _{max} at 280	0.9
6	Abutilon indicum, Malvaceae	Quercetin	RP-HPLC	Solvent A - 0.5% aqeuous solution of Orthophosphoric acid Solvent B -methanol	C-18 ODS	Tunable absorbance λ _{max} 254	1.0
7	Aesculus hippocastanum, Hippocastanaceae	Esculin and fraxin	HPLC	Solvent A- acetic acid (1%) Solvent B- methanol Ratio: 84:16 v/v	RP 18	UV λ_{max} at 340	1.0
8	Alpinia nigra, Zingiberaceae	Astragalin and kaempferol-3- <i>O</i> - glucuronide	HPLC	Solvent A-acetonitrile Solvent B- 0.1 % aqueous acetic acid Ratio: 20: 80	RP 18	UV λ _{max} at 267	1.0
9	Lonicera edulis, Caprifoliaceae	4-aminobenzoic acid, rutin, quercitrin, gallic acid	HPLC-ED	Solvent A- methanol 60 % Solvent B- 0.065 M acetic acid 40 % (y/y)	RP C- 18	Electrochemic al detector	1.0
10	<i>Michelia alba,</i> Magnoliaceae	Gallic acid, catechin, rutin ellagic acid and quercetin	HPLC	Solvent A- water- acetic acid (25:1 v/v) Solvent B- methanol	RP C- 18	UV λ _{max} at 280	1.0
11	<i>Vitis</i> sp. Vitaceae	Anthocyanins	HPLC	Solvent A-1.5% phosphoric acid Solvent B- 1.5% phosphoric acid, 20% acetic acid 25% acetonitrile	C-18	Diode array λ_{max} at 530	1.0
12	Uncaria tomentosa, Rubiaceae	Indole alkaloids	HPLC-MS	Solvent A- 30 mili molar ammonium acetate at pH 5 Solvent B- methanol:acetonitrile Ratio: 1:1	RP C- 18	Photodiode array	1.0
13	Terminalia arjuna, Combretaceae	Arjunic acid, arjunolic acid, arjungenin, arjunetin	HPLC	Acetonitrile:water Ratio: 30:70	ODS2 (RP)	UV-VIS λ_{max} at 220	0.8

14	Spiraea thunbergii, Rosaceae	Tulipalin B	HPLC	6% ACN in H ₂ O	C-18	Photodiode array	0.1
15	<i>Glycine max</i> , Fabaceae	Sphingolipids	HPLC	Solvent A- hexane Solvent B - 2-propanol/ethyl acetate/ 88% formic acid Ratio: 50:50:0.5 v/v	Si60	HPLC- ELSD	0.8
16	Salvia divinorum, Lamiaceae	Salvinorin A and B	HPLC	Acetonitrile: water Ratio: 1:1 v/v	C-18	UV λ _{max} at 210	0.1
17	Rubus glaucus, R. adenotrichus, Rosaceae	Ellagitannins, anthocyanins	HPLC/DAD	Solvent A- 2% aqueous formic acid Solvent B- acetonitrile/water/formic acid Ratio: 80:18:2 v/v/v	ODS-2	Diode array λ_{max} at 200 and 600	0.5
18	Pseudolarix kaempferi, Pinaceae	Pseudolaric acids A and B	HPLC	Solvent A- acetonitrile Solvent B-1% aqueous acetic acid, in which acetonitrile was linearly changed from 30% to 60% in 30 min	RP-C18	UV λ_{max} at 260	0.1
19	Prunus serotina, Rosaceae,	Flavonoid	RP-HPLC	Solvent A- 0.5% aqueous solution of orthophosphoric acid Solvent B-MeOH	ODS- C-18	UV–VIS λ_{max} at 370	2.0
20	Prunus cerasus, Rosaceae	Melatonin	HPLC	0.1 M potassium phosphate buffer with acetonitrile (20%)	OD53	Eight- channel Coul array	0.1
21	Phyllanthus urinaria, P. virgatus, P.maderaspatensis P. debilis Euphorbiaceae	Phyllanthin and hypophyllanthin	HPLC	Methanol:water Ratio: 70:30	S10 ODS2	Photodiode array λ _{max} at 220	0.7
22	Passiflora caerulea, P. incarnate, Passifloraceae	Flavonoids	HPLC	Methanol water Ratio: 1:1	C-18	UV-VIS λ_{max} at 340	1.0
23	Sanguinaria canadensis , Dicranostigma lactucoides, Papaveraceae	Benzo[c] phenanthridine alkaloids	HPLC	Solvent A- 25% acetonitrile Solvent B- 60% acetonitrile (v/v)	C-12	Diode array λ _{max} at 180– 550	0.5
24	<i>Opuntia</i> sp., Cactaceae	Betaxanthin and betacyanin	HPLC, HPLC-MS	Solvent A- water Solvent B- methanol	RP C- 18	Diode array λ_{max} at 482 and 535	1.0
25	Olea europea, Oleaceae	Organic acids (oxalic, citric, malic and succinic)	HPLC	0.1% (w/v) phosphoric acid in distilled water	KC-118	UV λ _{max} at 214	0.8
26	Ocimum spp. Lamiaceae	Ursolic acid	HPLC	Solvent A- acetonitrile Solvent B- 1.25% H ₃ PO ₄ Ratio: 86:14 (v/v)	RP- ODS	Photodiode array λ _{max} at 206	0.5

						VOIC	
27	Nothapodytes foetida, Icacinaceae	Camptothecinoids	HPLC	Acetonitrile/H2O Ratio: 25:75 v/v	C-18	UV λ _{max} at 970	1.0
28	Mitracarpus scaber, Rubiaceae	Triterpenic acids	HPLC	Acetonitrile/H2O Ratio: 85:15 v/v	ODC	UV λ _{max} at 215	0.6
29	Mammea americana, Clusiaceae	Coumarins, mammea	HPLC	H ₂ O:TFA:MeOH Ratio: 15:0.05:84.95 v/v/v	C-18	UV λ _{max} at 290	0.8
30	Solanum lycopersicum, Solanaceae	Lycopene	RP- HPLC	Solvent A- acetonitrile Solvent B- 1:1:1 mixture of methanol, hexane and methylene chloride	C-18	Diode array λ_{max} at 471	0.8
31	<i>Gmelina arborea,</i> Verbenaceae	Apigenin	RP- HPLC	Acetonitrile and distilled water in volume ratio of 45:55	ODS- 3V-C18	UV-970 λ _{max} at 340	1.0
32	Garcinia parvifolia, Guttiferae	Mangostin	RP- HPLC	Methanol and water Ratio: 95:5 %v/v	RP C-18	UV A _{max} at 319	1.0
33	Cassia alata, Leguminosae	Kaempferol-3-0- gentiobioside	HPLC	Solvent A- 1.25% aq AcOHy Solvent B- MeCN Ratio: 4:1	SPD	ODS-A 349	0.8
34	Alpinia nigra, Zingiberaceae	Kaempferol-3-0- glucuronide	HPLC	Solvent A- acetonitrile Solvent B- 0.1 % aqueous acetic acid Ratio: 20: 80	RP C-18	UV λ_{max} at 267	1.0
35	Caesalpinia pulcherrima, Caesalpiniaceae	Gallic acid, catechin, rutin, ellagic acid and quercetin	HPLC	Water-acetic acid Ratio: 25:1 v/v	RP C-18	UV λ _{max} at 280	1.0
36	Diospyros kaki, Ebenaceae	Barbinervic acid, Rotungenic acid	HPLC	Solvent A- methanol Solvent B - 0.1% aqueous H ₃ PO ₄ Ratio: 80:20 v/v	SB- C- 18	UV 210	1.0
37	Chrysophyllum roxburghii, Sapotaceae	Carotenoids	HPLC	Acetonitrile, methanol and ethyl acetate containing 0.05% of triethylamine	C-18	Photodiode array λ _{max} at 300- 600	0.5
38	Eucalyptus polyanthemos, Myrtaceae	Sideroxylonals	RP- HPLC	Acetonitrile: water containing 0.1% trifluroacetic acid	C-18 RS	Photodiode array λ _{max} at 275	0.75
39	Malus pumila, Rosaceae	Anthocyanin	HPLC	Water : methanol	C-18	Photodiode array λ _{max} at 520	1.0
40	Euphoria longana, Sapindaceae	Gallic and ellagic acid	HPLC	Solvent A- 0.4% formic acid Solvent B- methanol	RP-18	UV λ _{max} at 270	1.0
41	Enterophospora infrequens, Entrophosporaceae	Camptothecin	RP- HPLC	Water and acetonitrile	RP-18	Diode array λ_{max} at 256	0.5

PHYTOCONSTITUENTS EVALUATED VIA HPLC[5-45]

Large number of phytoconstituents were quantified via HPLC technique for example: flavonoids (quercetin, kaempferol, cyanidin); flavonoid glycosides (rutin); tannins (ellagic acid, ellagitannins); alkaloids (camptothecin, phyllanthin); polyphenols (arbutin); coumarins; triterpenes (ursolic acid, arjunolic acid); carotenoids (lycopene); sphingolipids etc. Some of reported quantified phytoconstituents were shown in figure 1.





Quercetin

Kaempferol

Cyanidin



Pelargonidin

Ellagitannins



'nн Coumaric acid



Ellagic acid



Arbutin

Esculin

Arjunic acid



Phyllanthin

Fraxin





Sphingolipids

Rutin



HPLC methods have been used in the determination of drugs in biological fluids and in pharmaceutical dosage forms. HPLC determination method with spectroscopic detection is useful for routine quality control of drugs in pharmaceutical formulations and stability studies. Various types of detectors: photo diode array, ultraviolet (UV), ultraviolet visible (UV-VIS), fluorometric, fluorescence have been employed along with different types of columns (C-8, RP-8, RP C-18, SB-CN, ODS-2) depending upon the synthetic drug to be evaluated. Some of reported synthetic drug molecules evaluated *via* HPLC were shown in table 2 given below.

Sr.no.	Drug	Drug category	Method	Mobile phase	Column	Detector and wavelength (nm)	Flow rate (mL/min)
1	Lamivudine, stavudine, nevirapine, zidovudine	Antiviral	RP-HPLC	Solvent A-50 mM NaH ₂ PO ₄ at pH 3.8 Solvent B- acetonitrile Ratio: 95:5 to 45:55 w/v	C-8	Photo diode array	0.5 to 1.0
2	4-bromo-2,5- dimethoxyphenethylamine	Hallucinogenic	HPLC	Acetonitrile/Potassium phosphate (pH 3.2)	RP-8	Diode array	1.0
3	Acetaminophen	NSAID	RP-HPLC	Mobile phase was prepared by adding 330 ml of methanol to 660 ml of the water pH 3.0 with 10% orthophosphoric acid	C-18	Photodiode array λ_{max} at 193.3	1.78
4	Amoxicillin	Antibiotic	RP-HPLC	Solvent A-95% phosphate buffer (0.01mol/L) pH4.8 Solvent B-5% acetonitrile mixture	RP C-18	UV229	1.3
5	Amphetamine	CNS stimulant	RP-HPLC	Solvent A-aqueous orthophosphoric acid (pH 2.1) Solvent B-acetonitrile Ratio: 90:10 v/v	C-8, C-18	Diode-array UV λ_{max} at 205	1.5
6	Atenolol	Beta blocker	RP-HPLC	Water: Buffer: Methanol Ratio: 50:35:15	Zorbax SB-CN	UV-VIS λ _{max} at 286	1.2
1	Hydrochlorothiazide	Diuretic	RP-HPLC	Water: Buffer: Methanol Ratio: 50:35:15	Zorbax SB-CN	UV-VIS λ _{max} at 286	1.2
8	Benzalkonium chloride	Preservative	HPLC	Solvent A- potassium dihydrogen orthophosphate butfer (pH 5.5) Solvent B- acetonitrile Ratio: 40:60 v/v	Waters Spherisor CN	UV λ_{max} at 210	1.0
9	Cephalosporins (Cefepime)	Antibiotic	HPLC	Solvent A- 90/10 (5 mM potassium phosphate/ Acetonitrile) Solvent B- 50/50 (5 mM potassium phosphate/ Acetonitrile)	C-18	UV λ _{max} at 254	0.2
10	Ceftriaxone	Antibiotic	RP-HPLC	Methanol:water:acetonitrile Ratio: 80:15:5 v/v/v	C-18	UV-VIS λ _{max} at 270	1.0
11	Tiaprofenic acid, flurbiprofen, diclofenac acid, mefenamic acid	NSAID	RP-HPLC	Methanol.water.acetonitrile Ratio: 80:15:5 v/v/v	C-18	UV-VIS λ _{max} at 270	1.0
12	Hydrocortisone, corticosterone	Corticosteroids	RP-HPLC	Solvent A- water Solvent B- 50% acetonitrile, 17% methanol, 33% isopropanol	C-18	Diode-array	1.0
13	Lercanidipine	Antianginal	RP-HPLC	Solvent A- acetonitrile and an aqueous solution of 1.5% triethylamine, pH adjusted to 3.0 Solvent B- orthophosphoric acid Ratio: 35:65	C-18	UV–VIS λ_{max} at 240	1.0

μ μ
Image: Section of the sectin of the section of the section of th
B Rămpkin Arhiniz PEIC Short A. 16 N. prasim dingtage G.8 Dikerung 12 1 Marine Marine Juin 1250 v Juin 1250 v Juin 1250 v Glikinik Anfoldzik BFUC Accumite durater C18 UV 12 1 Marine Marine Silor 4- Marinik BFUC Silor 4- Marinik Glikinik Anfoldzik BFUC Accumite durater C18 UV Juin 120 1 Pasterul, optimization Silor 4- Marinik BFUC Marinik BFUC Marinik durater C18 UV Juin 120 10 Pasterul, optimization Silor 4- Marinik durater C18 UV Silor 4- Marinik durater C18 UV Juin 120 10 Pasterul Silor 4- Marinik durater Ling 120 Lin
No.
Image: Solution of the security of the
Image: space
b Phychkam, upphrhame NSB IIIC North-Actaritie OS2 U/H II 10 Phychkam, upphrhame NSB IIIC North-Actaritie 062 U/H 1 Ampychic HILC Mainell North-Actaritie 1 Ampychic HILC Mainell North-Actaritie 1 Ampychic HILC Mainell U/H 0 <td< td=""></td<>
B Automation Note
1 Symposition 1 Symposition 1 $\frac{1}{2}$ maxmal, schninghet, fyrore and (2) : 3: 3: Num HEC 0.00000000000000000000000000000000000
10 rankana, soluti Number for how NM programmation diputes how NM programmation diputes how NM programmatic sprogram how NM programmatic sprogrammatic sprogram how NM programmatic sprogrammatic sprogrammatr
adminiput uppor adminiput uppor $\lambda_m \approx 0.2$ </td
Image: Indication of the X, N, N Image: Indication of the X, N </td
18 Catara Obsiminal HPLC UNI M MPL (actimitic properties during properis during properimentis during properis during properties during p
δ δ
Image: control in the start 40 in the
19 Nediscain Antibitic RP-RPLC Solies 14-20 mM sodim hydrogen phosphate C18 U/V18 - λ _m at 200 10 Nenuscrine Antibitic Ratic: 60-40 v/v C18 Numeration Numeration Ratic: 60-40 v/v C18 Fluctorentric 1.0 10 Nimuzepine Antilepressan RP-RPLC Phosphate buffer: Activitile Ratic: 718/V C18 UV 1.0 Antilepressan Pherplate buffer: Activitile Ratic: 718/V C18 UV 1.0 Antilepressan Pherplate buffer: Activitile Ratic: 718/V C18 UV 1.0 Antilepressan Pherplate buffer: Activitile Ratic: 718/V C18 UV 1.0 11 Mitracipan Antilepressan RP-HPLC Phosphate buffer: Activitile Ratic: 718/V C18 UV 1.0 Numeration Antilepressan RP-HPLC Phosphate buffer: (014) methanol Ratic: 718/V Ratic: 718/V C18 V/v 1.0 12 Methoteant Anticaner HPLC Phosphate buffer: (014) methanol Ratic: 718/V RP-HPLC Nomatainol Ratic: 718/V C18
Anison Main difference
Nimusepine Antidepresant RP-HPLC Nomeschnichte (M-M) L <thl< th=""> L<</thl<>
Image: Constraint of the spectral product with the spectral produ
D Nitrozzejne Anidepresant RP-HPLC Phosphate buffer: Acetoninile C-18 UV 1.0 hypertensive Minaciprant Anidepresant RP-HPLC Phosphate buffer: Acetoninile C-18 UV 1.0 38 Trimethoprim Antibiotic HPLC Minaciprant Antice of acconinitile and 0.5% triethylamine in No.05% C-18 UV 1.5 21 Minacipran Anidepresant RP-HPLC Phosphate buffer (pH-4): mehanol manufer in the fill (pH-4):
Image: Constraint of the sector in the s
21 Milnacipran Antidepressant RP-HPLC Phosphate buffer activityile Radio: 72.28 v/v C-18 UV and spectrofluorometric λ _{mat} at 220 10 Phosphate buffer (pH 4): mehanol Radio: 18.82 v/v 10 Name at 221 22 Mehotrexate Anticancer HPLC Phosphate-tirate buffer (pH 4): mehanol Radio: 30:70 v/v RP 4 UV - 23 Mehotrexate NSAID HPLC Solvent A: Phosphate buffer (02.N) Solvent B: activityile Radio: 38.62 v/v RP C18 UV 0.5 -
Ratio: Ratio: 7.28 v/v spectrofluorometric λ _{mat} dt 220 Ratio: Ratio: Ratio: Ratio: Ratio: No. N
Image: Constraint of the sector infinite relation of the secto
22 Methotexate Anticancer HPLC Phosphate-citrate buffer (βH 4): methanol Ratio: 30:70 v/v RP-8 UV \lambda_mat 432 39 Zidovudine Anti-hiv RP-HPLC Orthophosphoric acid : acetonitrile Ratio: 73:27 v/v C-18 UV, photo diode array \lambda_max at 246 0.9 23 Meloxicam NSAID HPLC Solvent A- Phosphate buffer (02.N) Ratio: 36:20 v/v RP -18 UV 0.5 40 Nicorandil Antianginal Antianginal HPLC Methanol and 0.002 mol/L phosphate buffer (pH 7.0) Ratio: 37 v/v ODS C-18 diode array \lambda_max at 240 1.0 24 Flutamide Antiandrogenic HPLC Actiontifictertan/drofugan/THF) water C-18 UV 1.0
Ratio: 30/10 v/v RP-8 UV λ _{mm} dt 342 23 Meloxicam NSAID HPLC Solvent A: Phosphate buffer (02.N) Solvent B: acetonitrile Ratio: 38/52 v/v RP C-18 UV 0.5 24 Flutamide Antiandrogenic HPLC Action/THF/water C-18 UV 0.5 24 Flutamide Antiandrogenic HPLC Action/ThF/water C-18 UV 1.0
Image: Solution in the sector in the sec
23 Meloxicam NSAID HPLC Solvent A- Phosphate buffer (0.2.N) Solvent B- acetointrile Ratio: 38/52 v/v RP C-18 UV 0.5 24 Flatamide Antiandrogenic HPLC Actiont/Tetranoffican/THF/water C-18 UV 0.5 24 Flatamide Antiandrogenic HPLC Action/Tetranoffican/THF/water C-18 UV 1.0
$\frac{1}{24} = \frac{1}{1} + 1$
24 Flatamide Antiandrogenic HPLC Acctonitrileteralvalorfuran/THF1xwater C-18 UV 1.0 41 Ambroxol Secretolytic HPLC Acctonitrile: methanol : 0.5% ammonium acetate C-18
24 Flutamide Anfandrogenic HPLC Acetonitrile: methanol : 0.5% ammonium acetate C-18
27 Indulitov Antoniovicul III Lo Avvolutinavicul III (vani
Perfor: 115/2566 UV 0.8
N Destinges hetworkscope Conjugation (d) LIDI C According for the second
2) Heatomic calculations, contractions in the Activities water in the Activities and Ammonium RP-C-18 UV-VIS 0.3
$\frac{1}{2} \sum_{n=1}^{n} \frac{1}{n} \sum_{n=1}^{n} \frac{1}$
20 Dexandratisene Unitionetitude Ke-HrLU Mediation: Valer U-18 UV 1.0 Ratio: 42:58
KRDC: N/20 Ameria 124 43 Sodium ozagrel Antiinflammat HPLC Sodium phosphate buffer (0.05 moll, pH 3.0) and C-18 Photodiode array 1.5
$\frac{21}{10}$ Chlorotrazane Skeletal KP-HPLC Accondition and builde distilled water RP4-8 UV 1.0 ory accondition λ_{max} at 276
muscle Ratio: 60:40 A _{mat} at 250 Ratio: 94:6 v/v for plasma
relatant Ratio: 96: 4 v/v for urine
28 Rakotifene Estrogen RP-HPLC Acetonitrile and phosphate buffer C-18 UV 1.0 44 Ketamine Veterinary HPLC Acetonitrile.0.03 mol/L phosphate buffer C-18 Photodiode array 1.5
receptor Ratio: 30:70 viv λ_{max} at 200 anesthetic pH (7.2) λ_{max} at 210
modulator Ratio: 23:77 viv

SYNTHETIC MOLECULES EVALUATED *VIA* HPLC [46-86]

Synthetic molecules were successfully evaluated both qualitatively as well as quantitatively via HPLC as shown in figure 2. Several pharmaceutically important Antiviral drugs: lamivudine, stavudine, nevirapine, zidovudine; NSAIDS: acetaminophen, meloxicam, paracetamol, phenylbutazone; Antibiotics: amoxicillin, cefepime, rifampicin; CNS stimulants: amphetamine; Beta blockers: atenolol; Preservatives: benzalkonium chloride; Diuretics: hydrochlorothiazide; Antidiabetics: metformin; Anticancer: prednisone, methotrexate; Corticosteroids: betamethasone. dexamethasone; Antianginal: Lercanidipine, Nicorandil; Immunomodulator : azathioprine: Anti-allergic: cetirizine: Antidepressant: Nitroxazepine, milnacipran; Veterinary anesthetic: Ketamine: Antiinflammatory: sodium ozagrel; Secretolytic: Ambroxol were successfully evaluated via HPLC.







Paracetamol

Rifampicin



Phenylbutazone

Caffeine



Ketamine







Norfloxacin

Metformin

Ambroxol

Lamivudine



Nevirapine







Zidovudine

Acetaminophen





Amphetamine







Benzalkonium chloride

Cefepime

Atenolol







Nicorandil

Alfuzosin



Methotrexate

Trimethoprim



Itraconazole





Diclofenac acid

Hydrocortisone

Olanzapine

Figure 2: Synthetic drug molecules evaluated via HPLC

Meloxicam





Flutamide

CONCLUSION

HPLC is one mode of chromatography, one of the most used analytical techniques. HPLC applications can be used effectively for screening analysis as well as quality evaluation of natural as well as synthetic compounds. Owing to the simplicity and efficiency of HPLC specific and rapid determination of various natural and synthetic compounds can be carried out. HPLC can be employed for the routine analysis of natural and synthetic compounds pharmaceutical in formulations and in bulk drug preparations as well as for the quality assurance of related extracts and market samples. Interest in HPLC has increased with improvements in its instrumentation and methods and especially in the last few years, with the combination of hyphenated techniques like HPLC-MS/MS, HPLC/DAD/MS, HPLC-ED.

REFERENCES

1).Ahuja S and Dong MW. Handbook of Pharmaceutical Analysis by HPLC. 1st ed. Academic Press Publisher.UK 2005.

2).Ahuja S and Alsante K. Handbook of Isolation and Characterization of Impurities in Pharmaceuticals. Academic Press New York 2003.

3).Snyder LR and Kirkland JJ. Introduction to Modern Liquid Chromatography. John Wiley and Sons New York 1979.

4).Meyer VR. Practical HPLC. Wiley New York 1998.

5).Correia H et al. Polyphenolic profile characterization of *Agrimonia eupatoria* L. by HPLC with different detection devices. Biomedical Chromatography 2006; 20: 88-94.

6). Aaby K et al. Characterization of Phenolic Compounds in Strawberry (*Fragaria* \times *ananassa*) Fruits by Different HPLC Detectors and Contribution of Individual compounds to Total Antioxidant Capacity. Journal of Agricultural and Food Chemistry 2007; 55: 4395-4406.

7).Pothitirat W and Gritsanapan W. HPLC Quantitative Analysis Method for the Determination of α -Mangostin in Mangosteen Fruit Rind Extract. Thai Journal of Agricultural Science 2009; 42(1): 7-12.

8).Mousavinejad G et al. Identification and quantification of phenolic compounds and their

effects on antioxidant activity in pomegranate juices of eight Iranian cultivars. Food Chemistry 2009; 115: 1274–1278.

9).Kittipongpatana N et al. High-Performance Liquid Chromatographic Method for Separation and Quantitative Analysis of Arbutin in Plant Tissue Cultures. CMU. Journal of Natural Science 2007; 6 (1): 65.

10).Rajalakshmi PV and Senthil KK. Direct HPLC analysis of quercetin in exudates of *Abutilon indicum* (Linn). Malvaceae. Journal of Pharmaceutical Science and Technology 2009: 1 (2): 80-83.

11).Stanic G et al. HPLC Analysis of Esculin and Fraxin in Horse-Chestnut Bark (*Aesculus hippocastanum* L.). Croatica Chemica Acta 1999; 72 (4): 827-834.

12).Qiao C et al. HPLC determination of two bioactive flavone glycosides and GC-MS analysis of volatile oil constituents in *Alpinia nigra*. Asian Journal of Traditional Medicines 2007; 2 (3): 85-91.

13).Gazdik Z et al. Use of Liquid Chromatography with Electrochemical Detection for the Determination of Antioxidants in Less Common Fruits. Molecules 2008; 13: 2823-2836.

14).Samee W and Vorarat S. Simultaneous Determination of Gallic acid, Catechin, Rutin, Ellagic Acid and Quercetin in Flower Extracts of *Michelia alba, Caesalpinia pulcherrima* and *Nelumbo nucifera* by HPLC. Journal of Thai Pharm Health Science; 2007: 2(2):131-137.

15).Liu X et al. Quantification and Purification of Mulberry Anthocyanins with Macroporous Resins. Journal of Biomedicine and Biotechnology 2004; 5 (2004) 326–331.

16).Montoro P et al. Identification and Quantification of Components in Extracts of *Uncaria tomentosa* by HPLC-ES/MS. Phytochemical analysis 2004; 15: 55-64.

17).Singh DV et al. RP-LC determination of oleane derivatives in *Terminalia arjuna*. Journal of Pharmaceutical and Biomedical Analysis 2002; 28: 447–452.

18).Lee J et al. Antibacterial effects of S-(-)-tulipalin B isolated from *Spiraea thunbergii* Sieb. on *Escherichia coli*, a major food borne pathogenic microorganism. Journal of Medicinal Plants Research 2008; 2(3): 059-065.

19).Wang L et al. HPLC Quantification of Sphingolipids in Soybeans with Modified Palmitate Content. Journal of Agriculture and Food Chemistry 2006.

20)Tsujikawa K et al. Determination of salvinorin A and salvinorin B in *Salvia divinorum*-related products circulated in Japan. Forensic Science International 2008; 180: 105–109. 21).Mertz C et al. Analysis of Phenolic Compounds in Two Blackberry Species (*Rubus glaucus* and *Rubus adenotrichus*) by High-Performance Liquid Chromatography with Diode Array Detection and Electrospray Ion Trap Mass Spectrometry.

Journal of Agriculture and Food Chemistry 2007; 55: 8616-8624.

22).Qiao CF. HPLC Analysis of Bioactive Diterpenoids from the Root Bark of Pseudolarix kaempferi. Journal of Food and Drug Analysis 2006; 14 (4): 353-356.

23).Olszewska M. Quantitative HPLC analysis of flavonoids and chlorogenic acid in the leaves and inflorescences of Prunus serotina. Acta chromatographica 2007; 19: 253-269.

24).Burkhardt S et al. Detection and Quantification of the Antioxidant Melatonin in Montmorency and Balaton Tart Cherries (Prunus cerasus). Journal of Agriculture and Food Chemistry 2001; 49: 4898-4902.

25). Tripathi AK et al. Quantitative Determination of Phyllanthin and Hypophyllanthin in Phyllanthus Species by High-performance Thin Layer Chromatography. Phytochemical Analysis 2006; 17: 394-397.

and Haustein C. 26).Frye Α Extraction. identification, and quantification of Harmala Alkaloids in Three Species of *Passiflora*. American Journal of Undergraduate Research 2007; 6 (3): 19-26.

27).Suchomelova J et al. HPLC quantification of seven quaternary benzo[c]phenanthridine alkaloids in six species of the family Papaveraceae. Journal of Pharmaceutical and Biomedical Analysis 2007; 44: 283–287.

28).Castellanos-Santiago E and Yahia EM. Identification and Quantification of Betalains from the Fruits of 10 Mexican Prickly Pear Cultivars by High-Performance Liquid Chromatography and Electrospray Ionization Mass Spectrometry. Journal of Agriculture and Food Chemistry 2008; 56: 5758-5764.

29).Ergonul PG and Nergiz C. Determination of Organic Acids in Olive Fruit by HPLC. Czech Journal of Food and Science 2010; 28 (3): 202-205. 30).Silva MGV et al. Variation of Ursolic Acid Content in Eight Ocimum Species from Northeastern Brazil. Molecules 2008; 13: 2482-2487.

31).Wu SF. et al. Camptothecinoids from the seeds of Taiwanese Nothapodytes foetida. Molecules 2008; 13: 1361-1371.

44).Rangkadilok N et al. Identification and

32).Gbaguidi F et al. HPLC quantification of two isomeric triterpenic acids isolated from Mitracarpus scaber and antimicrobial activity on *Dermatophilus congolensis*. Journal of Pharmaceutical and Biomedical Analysis 2005; 39: 990-995.

33).Perez OP et al. Isolation and characterisation of active compounds from Mammea americana Lin. Revista cubanade quimica 2007; 19 (1): 74-77.

34).Bicanic D et al. Quantification of lycopene in tomato products: comparing the performances of a newly proposed direct photothermal method and high-performance liquid chromatography. Journal of the Science of Food and Agriculture 2005; 85: 1149-1153.

35). Adhyapak S. High Performance Liquid Chromatographic method for quantization of apigenin from dried root powder of Gmelina arborea Linn. International Journal of Pharma and Bio Sciences 2011; 2(1): 742-749.

36).Syamsudin et al. HPLC Analysis and pharmacokinetic study of Mangostin after orally administration in rats. The Pharma Research 2009; 2: 43-49.

37). Moriyama H et al. HPLC quantification of kaempferol-3-Ogentiobioside in *Cassia alata*. Fitoterapia 2003; 74: 425-430.

38).Qiao C et al. HPLC determination of two bioactive flavone glycosides and GC-MS analysis of volatile oil constituents in *Alpinia nigra*. Asian Journal of Traditional Medicines 2007; 2 (3): 85-91.

39).Samee W and Vorarat S. Simultaneous Determination of Gallic acid, Catechin, Rutin, Ellagic Acid and Quercetin in Flower Extracts of Michelia alba, Caesalpinia pulcherrima and Nelumbo nucifera by HPLC. Journal of Thai Pharm Health Science 2007; 2(2): 131-137.

40). Fan JP and He CH. Simultaneous quantification of three major bioactive triterpene acids in the leaves of *Diospyros* kaki by high-performance liquid chromatography method. Journal of Pharmaceutical and Biomedical Analysis 2006; 41:950-956.

41).Chandrika UG al. Identification et and hplc quantification of carotenoids of the fruit pulp of Chrysophyllum roxburghii. Journal of National Science Foundation of Sri Lanka 20005; 33 (2): 93-98.

42).Wallis IR et al. Quantification of sideroxylonals in Eucalyptus Foliage by High performance Liquid Chromatograpy. Phytochemical Analysis 2003; 14: 360-365.

43).Mulabagal V. et al. Cultivars of Apple Fruits That Are Not Marketed with Potential for Anthocyanin Production. Journal of Agriculture and Food Chemistry 2007; 55: 8165-8169.

Quantification of Polyphenolic Compounds in Longan (Euphoria longana Lam.) Fruit. Faculty of Science,

Mahidol University 2004.

45).Amna T et al. Determination and quantification of camptothecin in an endophytic fungus by liquid chromatography – positive mode electrospray ionization tandem mass spectrometry. Current science 2006; 91 (2): 208-212.

46).Schuman M et al. HPLC analysis of generic antiretroviral drugs purchased in Rwanda. Bull. Soc. Sci. Med. 2005; 3: 317-324.

47).Cole MD et al. 4-Bromo-2, 5dimethoxyphenethylamine (2C-B): A review of the public domain literature. Science and Justice 2002; 42(4): 223-224.

48).Suzen S et al. Quantification of Acetaminophen in Pharmaceutical Formulations Using High Performance Liquid Chromatography. J. Fac. Pharm. Ankara 1998; 27(2): 93-100.

49).Pires LR et al. HPLC determination of amoxicillin comparative Bioavailability in healthy volunteers after a single dose administration. Journal of Pharmaceutical Science 2003; 6(2): 223-230.

50).Pavlova V and Jovanovic SP. Simultaneous Determination of Amphetamine, Methamphetamine and Caffeine in Seized Tablets by High Performance Liquid Chromatography. Acta Chromatographia 2007; 18: 157-167.

51).Zaveri M and Khandhar A. Development and Validation of a RP-HPLC for the Simultaneous Estimation of Atenolol and Hydrochlorothiazide in Pharmaceutical Dosage Forms. JPRHC; 2(3): 248-252.

52).Mehta J et al. Development and Validation of a Precise Method for Determination of Benzalkonium Chloride (BKC) Preservative in Pharmaceutical Formulation of Latanoprost Eye Drops. E-Journal of Chemistry 2010; 7(1): 11-20.

53).Hurum et al. Determination of Cefepime and Related Compounds Using HPLC with UV Detection. LC GC Asia Pacific 2009; 121(2).

54).Sultana N et al. Simultaneous Determination of Ceftriaxene Sodium and Non Steroidal Antiinflammatory Drugs in Pharmaceutical.

Formulations and Human Serum by RP-HPLC. Journal of the Chinese Chemical Society 2010; 57: 1278-1285

55).Woodwad C and Major R. Preparative Purification of Corticosteroids by HPLC; Scalability and Load ability using Agilent Prep C18 HPLC Columns. Agilent Technologies, Publication USA 2005.

56).Popovic I et al. LC Determination of Lercanidipine and its Impurities Using Drylab Software and Experimental Design Procedures. Chromatographia 2008; 67: 449-454.

57).Basavaiah K et al. Quantitative determination of Olanzapine in Pharmaceutical Preparations by HPLC. J.Mex. Chem. Soc. 2008; 52(2): 120-124.

58).Kumar AKH et al. A Validated High Performance liquid Chromatography Method for the Determination of rifampicin and desacetyl rifampicin in plasma and urine. Indian Journal Pharmacology 2004; 36(4): 231-233.

59).Jedziniak P et al. Determination of Phenylbutazone and Oxyphenbutazone in Bovine Plasma Using High Performance Liquid Chromatography with UV Detection. Bull Vet Inst Pulawy 2005; 49: 223-226.

60).Altun ML. HPLC Method for the Analysis of Paracetamol, Caffeine and Dipyrone. Turk J Chem 2002; 26: 521-528.

61).Bedor DCG et al. Development and Validation of a new method for the quantification of Norflpxacin by HPLC-UV and its application to a comparative pharmacokinetic study in human volunteers. Brazilian Journal of Pharmaceutical Sciences 2007; 43(2): 231-237.

62).Parveen PS et al. RP-HPLC Method for the Estimation of Nitroxazepine Hydrochloride in Pharmaceutical Dosage Forms. International Journal of Applied Biology and Pharmaceutical Technology 2010; 1(3): 1181-1187.

63).Mehta PJ and Khatri DM. Development and Validation of RP-HPLC Method for Determination of Milnacipran Hydrochloride in Pharmaceutical Formulations. International Journal of Pharmacy and Pharmaceutical Sciences 2010; 2: 137-141.

64).Yousefi G et al. Quantification of Polyethylene Glycol esters of Methotrexate and Determination of their Partition Coefficients by Validated HPLC Methods. Iranian Journal of Pharmaceutical Research 2009; 8(1): 27-31.

65).Mehmood KT et al. Specific and Simple HPLC assay of Ecofriendly Meloxicam in Pharmaceutical Formulations. Journal of Pharmaceutical Sciences and Research 2010; 2: 878-883.

66).Jalalizadeh H et al. A Rapid and Sensitive HPLC method for Determination of 2-Hydroxyflutamide in Human Plasma. International Journal of Pharmacology 2006; 2(2): 221-225.

67).Sangiorgi E and Curatolo M. Rapid Determination of Six Synthetic Corticosteroids in Urine by High Performance Liquid Chromatograph Atmospheric Pressure Chemical Ionisation Mass Spectometry. Istituto zooprofilattico sperimentale delle venezie: 969-975.

68).Kwak HW and Amico DJD. Determination of

Dexamethasone sodium phosphate In the vitreous by High Performance Liquid Chromatography. Korcan J. Opthalmol 1995; 9: 79-83.

69).Yadav AH et al. RP-HPLC method for Determination of Aceclofenac, Chloroxapine and Paracetamol in Bulk and Pharmaceutical Formulation. International Journal of Pharmaceutical Research and Development 2009; 1(10): 1-7.

70).Reddy PV et al. RP-HPLC Determination of Raloxifene in Pharmaceutical Tablets. Journal of Chemistry 2006; 3(10): 60-64.

71).Bhaskar M et al. RP-HPLC method Development for the Determination of Azathioprine in Bulk drug and Pharmaceutical Dosage Forms. International Journal of Chem.Tech Research 2010; 2(2): 1176-1179.

72).Yilmaz B. Reverse Phase HPLC Method for Determination of Nebivolol in Pharmaceutical Preparations. International Journal of Pharmaceutical Science Review and Research 2010; 1(2): 14-17.

73).Yang JF et al. Determination of Gliclazide in Human Plasma by High Performance Liquid Chromatography. Asian Journal of Drug Metabolism and Pharmacokinetics 2004; 4(3): 231-234.

74).Arayne MS et al. Determination and Quantification of Cetrizine HCl in Dosage Formulations by RP-HPLC. Pakistan Journal of Pharmaceutical Sciences 2005; 18(3): 7-11.

75).Barrett B et al. Validated HPLC-MS/MS Method for Determination of Quetiapine in human Plasma. Journal of Pharmaceutical and Biomedical Analysis 2007; 44: 498-505.

76).Chitlange SS et al. Stability Indicating RP-HPLC Method for Simultaneous Estimation of Valsartan and Amlodipine in Capsule Formulation. Asian J. Research Chem. 2008; 1(1): 15-18.

77).Reddy PB et al. Determination of Pantroprazole sodium and Lansoprazole Individual Dosage Form Tablets by RP-HPLC using Single Mobile Phase. International Journal of Applied Biology and Pharmaceutical Technology 2010; 1(2): 0976-4550.

78).Redmann S and Charles BG. A rapid HPLC Method with fluorometric detection for determination of plasma itraconazole and hydroxy-itraconazole concentration in cystic fibrosis children with allergic bronchopulmonary aspergillosis. Biomedical Chromatography 2005.

79).Shakya AK et al. Simple and Rapid HPLC Method for the Determination of Alfuzosin in Human Plasma. Jourdan Journal of Pharmaceutical Sciences 2010; 3(1): 25-35.

80).Lakkanatinaporn P and Matayatsuk C. Simultaneous HPLC method for Determination of Sodium Trimethoprim Phenylpropanol disulphonate and Sodium sulphaquinoxaline in veterinary drugs. Songklanakarin J. Sci. Tecnol. 2004; 26(6): 849-854.

81).Kabra B et al. Simultaneous Quantitative Determination of Zidovudine and nevirapine in human plasma using isocratic, reverse phase high performance liquid chromatography. Tropical Journal of Pharmaceutical Research 2009; 8(1): 79-86.

82).Cheng ZZ et al. Identification and Determination of Nicorandil and its Degradation Products by HPLC and GC/MS. Chinese Chemical Letters 2006; 17(8): 1057-1060.
83).Pasha K et al. RP-HPLC method for determination of Ambroxol in Pharmaceutical dosage forms. Research Journal of Pharmaceutical, Biological and Chemical Sciences 2010; 1(3): 250.

84).Lakshmi KS et al. Simultaneous Determination of Metformin and Pioglitazone by Reversed Phase HPLC in Pharmaceutical Dosage Forms. International Journal of Pharmacy and Pharmaceutical Sciences 2009; 1(2): 162-166. 85).Sun K et al. Determination of Sodium ozagrel in human plasma and urine by HPLC with solid- phase extraction and its application to pharmacokinetics. Asian Journal of Pharmaceutical Sciences 2006; 1(3-4): 222-228.

86).Bolze S and Boulieu R. HPLC determination of ketamine, norketamine and Dehydronorketamine in plasma with a high purity reversed- phase sorbent. Clinical Chemistry; 1998: 560-564.