

## MITOGEN ACTIVATED PROTEIN KINASE AT THE CROSSROADS OF ALZHEIMER'S DISEASES

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### Abstract

Alzheimer's disease (AD) is associated with various neurodegenerative alterations and inflammation thought to play a major role in its pathogenesis. The Mammalian stress activated protein kinase (SAPK), p38 mitogen-activated protein kinases (MAPKs), a family of serine/threonine protein kinases, activated in response to wide range of cellular stresses as well as in response to inflammatory mediators. A large body of evidences indicates that p38MAPK activity is critical for normal immune and inflammatory response. Moreover, the p38 MAPK pathway is considered to be a key regulator of various inflammatory pathways which are activated during normal aging and AD therapy. The p38MAPK pathway which is a key regulator of pro-inflammatory cytokines biosynthesis at the transcriptional and translational levels, which makes different components of this pathway, a potential targets for the treatment of autoimmune and inflammatory diseases. Furthermore, p38 MAPK is over expressed in AD and have been linked to A $\beta$  deposition and Tau tangle formation. Favourable modulator of p38 MAPK found to beneficial in a variety of experimental models of AD, further implicating p38 MAPK in AD pathogenesis. In this review, we provide an overview on p38 MAPK and its implication in the pathogenesis of Alzheimer's diseases.

**Key words:** - SB203580; p38 MAPK; Alzheimer's disease; ICV STZ; Memory; Oxidative stress.

### Introduction

Alzheimer's diseases (AD) is an age related irreversible, progressive neurodegenerative disorder characterized by amyloid beta deposition and Tau tangle formation and inability to form new memories and access existing ones, due to neuronal cell death in the hippocampus and frontal cortex [1,2]. Extensive evidences suggests that oxidative stress and neuroinflammation are the earlier events in AD pathology [3,4]. Moreover, oxidative stress and neuroinflammatory events have been linked to A $\beta$  deposition and Tau tangle formation [5]. Furthermore, during aging energy failure, oxidative stress and early increase in inflammatory mediators provides the basis for persistent stress to the neurons contributes to impaired synaptic functions, cognitive decline and neuronal death [6]. Although the neuropathological features of AD have been well defined, the underlying mechanisms responsible for the pathogenic processes have not been clearly delineated. This lack of understanding of the fundamental processes that are responsible for the neurodegeneration in AD likely is the reason there are no effective treatments to prevent the onset progression of the disease. However, research advances over the past several years have begun to provide some insight into the molecular mechanisms of AD. One particularly important area of investigation is the contribution of aberrant cell signaling events to the pathogenic process. For example, recent findings have provided strong evidence that the p38 mitogen activated protein (MAP) kinase signaling cascade is one signaling

pathway that overactivated in AD. Advances in molecular biology have led us to understand the molecular mechanisms involved in mediating stress response and subsequent neuronal death. Recently, it has been reported that stress activated protein kinases plays a major role in degeneration of neurons in AD pathology. Mainly, two SAPK's such as JNK and p38 MAPK have been implicated in tau hyperphosphorylation, amyloid beta deposition and in progression of inflammation that are being actively involved in the pathogenesis of AD [7-10] and may be attractive targets for neurodegenerative diseases [11,12]. In this review, we mainly focused on p38 mitogen activated protein kinase (MAPK) pathway, a major proinflammatory signal transduction pathway activated by various extracellular stimulation such as growth factors, oxidative stress, ultraviolet and cytokines, are hyperactivated in human AD brain [13-15]. *In vitro* and *In vivo* studies have been demonstrated that upon activation, p38 MAPK cascade leading to cause neuronal death through various pathways in brain including, activation of microglia/astroglia in the production of inflammatory mediators such as IL-1 $\beta$ , TNF- $\alpha$ , increase expression of COX/LOX and iNOS [16,17]. Oxidative/nitrosative stress and excitotoxicity which also contribute to phosphorylate p38 MAPK [15,18,19]. However, extracellular accumulation of A $\beta$  [7,8,10,20,21] and hyperphosphorylation of Tau protein [10,22-25] which are main hallmark of AD, are also activated via the phosphorylation of p38 MAPK in the hippocampal area of brain. Moreover, in Alzheimer's diseases, p38 MAPK also leads to desensitization of the insulin receptor [26,27] and degeneration of the cholinergic neurons, further leads to memory and cognitive dysfunction [20]. Neuronal apoptosis process also mediated by p38 MAPK [28]. Phosphorylated form of p38 MAPK also activate transcriptional factors such as MAPKAK2, PRAK, ATF-2, CHOP, NF-kB, AP-1, ATF-2 which further increases the expression of iNOS and many proinflammatory cytokines [29,30], all these changes are nearly relevant to the pathogenesis of AD brain via activation

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of p38 mitogen activated protein kinase and this hyperactivation of the p38 kinase and related signal pathway provides a new concept for discussing neuroinflammation in the aging human brain and may indicate a novel therapeutic target of the AD brain.

### Overview Of Mitogen activated protein kinase (MAPK)

Cellular behavior in response to extracellular stimuli is mediated through intracellular signaling pathways such as the mitogen-activated protein (MAP) kinase pathways [31]. MAP

**Table 1: p38 MAPK nomenclature and localization**

Name	Alternate name	localization
P38 $\alpha$	Sapk2b, csbp1	Ubiquitously
p38 $\beta$	Sapk2b, p38-2	Ubiquitously
p38 $\gamma$	Sapk3 erk6	Skeletal muscle
p38 $\delta$	Sapk4	Low in most tissues

**Table 2: Extracellular stimulants for the activation of p38 MAPK**

Growth factors	angiotensin ii, fgf, pdgf, vegf, tgf- $\beta$ , igf
Cytokines	Tnf- $\alpha$ , il-1 $\beta$ , cd154, il-17
Stress factors	Lps, mechanical stress, uv radiation, osmotic stress
Others	Thrombin, high glucose, endothelin, no, estradiol, insulin, gpcr, tkr

**Table 3: Enzymes and substrates for the activation and inactivation of p38 MAPK**

Upstream activator	Mapkkk ( mkkk or mekk 1/4, raf, ask1, mlk3, tak1, tao-1/3 ) mapkk( mkk3/4/6, mek1/2 )
Inactivation enzymes	Map ( mitogen activated protein kinase phosphatase) Serine/threonine protein phosphatase type 2c ( pp2c )
Downstream substrate	Mapkap2/mk2, mk3, lsp1, creb, elf-4e, mnk1/2, Prak
Transcriptional factors	Atf-1/2/6, srf, sap1, chop, p53, c/ebp $\beta$ , mef2a/2c, mitf1, ddit3, elk1, nfat, hbp1, ap-1, nf-kb, stat1/3

### p38 MAP Kinase

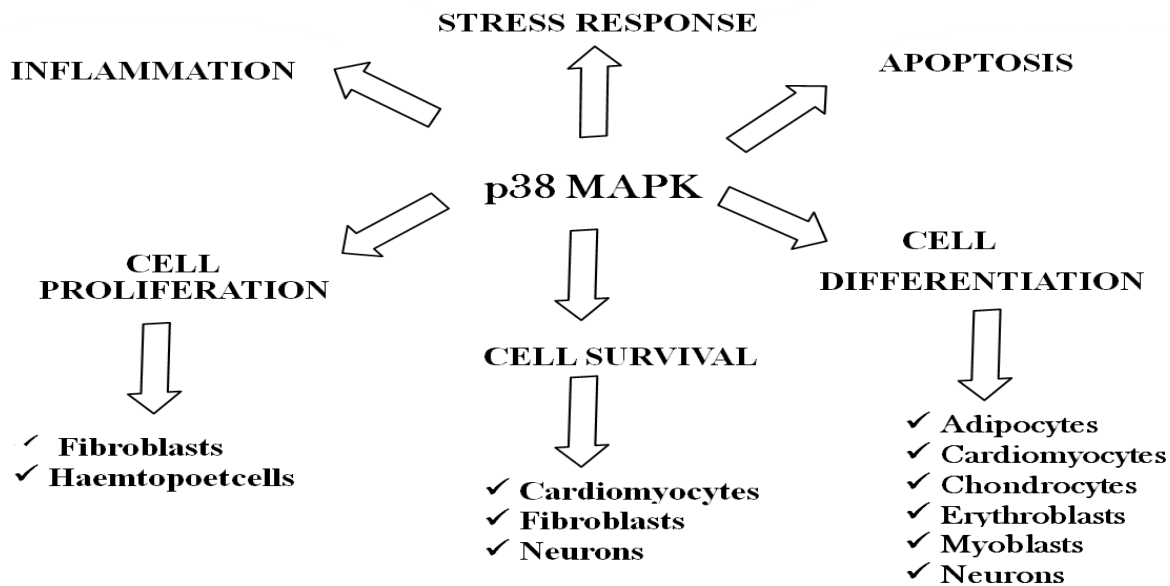
p38 $\alpha$  (p38) was first isolated as a 38 kDa protein and is known to be rapidly tyrosine phosphorylated in response to LPS stimulation [33]. p38 cDNA was also cloned as a molecule that binds puridiny l imidazole derivatives which are known to inhibit biosynthesis of inflammatory cytokines such as interleukin-1 (IL-1) and tumor-necrosis factor (TNF) in LPS stimulated monocytes [34]. To date, four splice variants of the p38 family have been identified: p38 $\alpha$ , p38 $\beta$  [35,36], p38 $\gamma$  [37,38], and p38 $\delta$  (SAPK4) [39]. Of these, p38 $\alpha$  and p38 $\beta$  are ubiquitously expressed while p38 $\gamma$  and p38 $\delta$  are differentially expressed depending on tissue type. All p38 kinases can be categorised by a Thr-Gly-Tyr (TGY) dual phosphorylation motif [40]. Sequence comparisons have revealed that each p38 isoform shares ~60% identity within the p38 group but only 40-45% to the other three MAP kinase family members [41].

### Localization of p38 MAPK in Brain

Despite an abundance of data concerning p38 activation and function in peripheral tissues, the role of p38 in the brain is poorly understood. This is surprising while considering the fact

kinases are members of discrete signaling cascades and serve as focal points in response to a variety of extracellular stimuli. Four distinct subgroups within the MAP kinase family have been described: (1) extracellular signal-regulated kinases (ERKs), (2) C-jun N-terminal or stress-activated protein kinases (JNK/SAPK), (3) ERK/ big MAP kinase-1 (BMK1), and (4) the p38 group of protein kinases. The focus of this review is to highlight the characteristics of the p38 kinases, components of this kinase cascade, activation of this pathway and the biological consequences of its activation [32].

that p38 is more highly expressed in brain than in peripheral tissues [35,39]. Only the p38 $\alpha$  and p38 $\beta$  isoforms are expressed in the brain, with high levels of protein in most major brain regions, including cerebral cortex, hippocampus, cerebellum, and several brainstem nuclei [29]. Detected mainly in neurons, p38 $\alpha$  is found in the nucleus, dendrites, and in cytoplasmic regions of the cell body. Both neurons and glia express p38 $\beta$ , with a pronounced nuclear location. The  $\alpha$  and  $\beta$  isoforms of p38 are especially enriched in hippocampus, the brain region predominantly involved in learning and memory. Further, they are heavily expressed in pyramidal neurons of CA1 and CA3 regions of hippocampus as well as in granule cells of the dentate gyrus [29,42]. Unlike p38 $\alpha$ , p38 $\beta$  is detected in glial cells of the CA1 region also. In addition to prominent mRNA and protein levels, p38 also exhibits a high basal activity in brain [29,43], which suggest that the p38 pathway may play a role in normal neuronal function in addition to its role as an SAPK. Unlike JNK, whose function(s) are preferentially related to the control of apoptosis, p38 in the brain is involved not only in apoptosis, but also in aspects of neuronal differentiation, synaptic function and neuronal plasticity. Additionally, the p38 pathway is active during the induction of



**Fig-1 Biological functions of p38 MAPK**

### Implication Of P38 Mapk In Alzheimer's Diseases At Various States

#### p38 MAPK Activation In Tau Hyperphosphorylation

p38 has been demonstrated in vitro to phosphorylate tau on residues known to be phosphorylated in NFTs extracted from AD brains [25,48-50]. However, many other kinases, such as glycogen synthase kinase (GSK)-3 $\beta$  and extracellular signal-regulated kinase (ERK)-2, have also been implicated as tau kinases, with different kinases demonstrating different preferences for certain sites [50]. Many of these kinases, including p38, have been demonstrated to phosphorylate tau when co-transfected with tau into cells, with GSK-3 $\beta$  demonstrating the best activity [51,52]. Although encouraging, the extrapolation of these data to AD is limited by caveats around overexpression and the use of non-neuronal cells. Therefore, current knowledge does not indicate the best target for intervention in tau phosphorylation in vivo, and further elucidation in this area is required. Additionally, it is not known if inhibition of any single kinase will have an impact on NFT formation. Although p38 has been implicated in tau phosphorylation in vitro and is associated in activated form with NFTs in AD in situ, there is currently no evidence that p38 phosphorylates tau in vivo. The activation of p38 has been demonstrated concurrently in conjunction with tau phosphorylation in the brains of rats implanted with IL-1-impregnated pellets; however, these results remain correlative [13,53]. Recently, p38 activation has been demonstrated in a murine amyloidogenic model of AD in which mutated APP is overexpressed, and activation correlated with plaque burden and tau phosphorylation in the brains of these mice [8]. Assessing a p38 inhibitor in such a model may elucidate the role of p38 in tau phosphorylation and plaque formation in vivo. However, it is important to keep in mind that although these mice appear to show some degree of tau hyperphosphorylation, they do not develop NFTs. Thus, the implications of decreasing tau phosphorylation on NFT formation for any therapeutic approach cannot be assessed in this model. Other transgenic mice, which overexpress tau alone or tau and APP, have been reported to form tangle-like structures and may therefore be useful tools for assessing potential therapeutics on NFT formation in vivo [54-56].

However, to date, the activation of kinases in these models has not yet been characterized.

#### p38 MAPK activation In Amyloid-beta

The in vitro effect of A $\beta$  on p38 in neurons is also controversial with a debate on whether A $\beta$  activates p38 in neurons. One research group has reported that A $\beta$  activates p38 in N2 neuroblastoma cells [7], while two other groups have not reported any such effect [57,58]. The recent finding that p38 is activated in double transgenic mice expressing ABPP (K670N/M671L) and PS1(P264L) [59], as determined by immunoblotting, has shown that A $\beta$  activates p38 in vivo. However, due to the lack of concurrent immunocytochemistry data, it is not clear whether this is microglial or neuronal in origin. Neuronal accumulation of A $\beta$  has been implicated as a cause of the neuronal loss that occurs in AD while previous studies have shown that A $\beta$  is cytotoxic to neurons, at least in cell culture. The mechanisms and signaling pathways involved are just beginning to be unraveled. It has been reported that A $\beta$  induces the activation of p38 in a concentration-dependent manner in both M17 human neuroblastoma cells and primary cortical neurons. Since A $\beta$  is present at up to micromolar concentration with the development of amyloid deposits, these findings suggest that the chronic exposure to high A $\beta$  levels may also be responsible for the abnormal activation of p38 in AD brain [60,61]. Some studies have demonstrated that inhibition of the p38 pathway either by overexpressing the dominant negative p38 or by specific pharmacological inhibitor such as SB203580 results in decreased A $\beta$ -mediated cytotoxicity indicating an important role for p38 in the toxication of A $\beta$ . However, since overexpression of dominant negative p38 or application of SB203580 does not completely block A $\beta$ -induced neuronal death, it is likely that p38 regulates neuronal death in concert with other signaling transduction pathways. In this regard, previous studies demonstrate that a related pathway, namely the JNK pathway, is also activated in cultured neuronal cells after exposure to A $\beta$  and that inhibition of JNK also partially attenuates A $\beta$ -induced cytotoxicity [57,58,62,63]. Therefore, it is conceivable that both the JNK and p38 pathways work synergistically in mediating A $\beta$ -induced neuronal death. In support of such a concept, another reports demonstrated a nearly complete overlap between phospho-JNK and phospho-p38 in severe AD cases implying

that JNK and p38 are both activated by the same signal and, as such, work synergistically in vivo [60,61]. It will now be of great interest to establish the mechanism by which A $\beta$  induces the activation of p38. In this regard, since A $\beta$  appears to bind to the surface of neurons through multiple receptors, and one or more of them may be involved in A $\beta$ -induced p38 activation, it is anticipated that the different receptors involved may lead to distinct responses. For example, the receptor for advanced glycation end products (RAGE), which interacts with A $\beta$  may be a good candidate that is specifically involved in mediating A $\beta$ -induced oxidative stress through the p38 pathway [64]. Also it is notable that A $\beta$  binding of A $\beta$ PP

induces A $\beta$ PP dimerization which, in turn, activates ASK1/MKK6/p38 cascade. It will also be important to characterize the mechanisms by which p38 activation leads to cell death. One possible target is TNF- $\alpha$ , because the expression of TNF- $\alpha$  is regulated by p38 [65]. More relevantly, the levels of both TNF- $\alpha$  are elevated in AD [66,67]. Therefore, the characterization of specific receptors as well as the downstream targets involved in A $\beta$ -mediated events will not only help to better understand the nature of A $\beta$  action but also identify specific targets for interrupting the pathogenesis process.

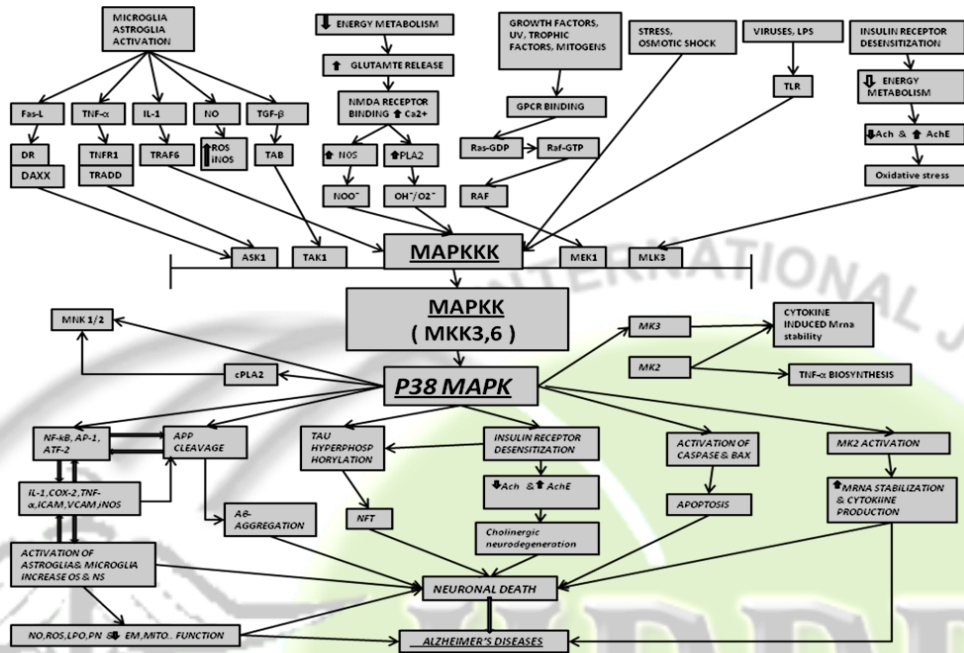


Fig- Neuronal death via activation of p38 MAPK

**p38 MAPK activation In Long Term Potentiation**

p38 MAPK has been reported to have higher expression in the hippocampus [29,68-70] and activation of p38 MAPK during stress conditions is reported to cause both short term and long term synaptic depression (LTD) [46,47,71-73]. But inhibition of p38 MAPK activity blocks the induction of LTD without affecting induction of LTP [42,46,70,74-77]. It has further been shown that there is complete absence or little formation of LTP induced by high frequency stimulation, whereas the low frequency stimulation enhances the formation of LTD in the hippocampus of STZ-induced diabetic rats [78-81].

**p38 MAPK activation In Neuroinflammation**

The activation of the p38 pathway plays essential role in the production of proinflammatory cytokines which has an integral role in Alzheimer’s disease development and may precede plaque and tangle formation [14,82]. Inflammatory components related to AD neuroinflammation include brain cells such as microglia and astrocytes and p38 MAPK is involved in the activation of microglia and astroglia cells known to generate beta-amyloid protein (A $\beta$ ) - one of the main pathologic features of AD [13,83]. Causative linkages between MAPK pathway activation and proinflammatory cytokine production by glia are based mainly on cell culture studies [18]. Phosphorylated p38 MAPK immunoreactivity was detected in microglia cells and co-localization with astrocytes in response to injury, illness, ageing or ischemia that begins a cascade of events which can be characterized as an inflammatory process [29,84]. In AD, the release of

proinflammatory cytokines and interleukins via activation of p38 MAPK is stimulated in activated microglia and astrocytes and these cells are characteristically found near damaged neurons and plaques [82,85,86]. Glia cells normally mediate the innate immune response in CNS, but when activated, they produce inflammatory mediators like cytokine S100 $\beta$ , IL-1, IL-6, TNF- $\alpha$ , prostaglandins and leukotriens, which increase the A $\beta$  activation. A $\beta$ , in turn, increases the levels of these cytokines which lead to plaque and tangle formation. The cytokines can simultaneously activate p38 MAPK to induce the generation of other mediators like nitric oxide (NO) toxic to neurons in AD brain [73,87-89]. It has also been demonstrated in in vitro models of inflammation that inducible COX-2 and inducible nitric oxide synthase expression requires the activation of p38 MAPK signaling pathway. A large number of research reports have provided evidences that support the involvement of inflammatory process as one of the key cascades in the development and worsening of Alzheimer’s disease in which p38 MAPK signaling pathway has been fully implicated [16,19].

**p38 MAPK activation In Oxidative Stress and Excitotoxicity**

Oxidative stress is the key feature for neuron cell death in AD [90](Butterfield et al., 2006) and p38 MAPK is highly sensitive to oxidative stress [91,92]. The oxidative stress induces several other key events like protein aggregation [93,94], mitochondrial dysfunction [95] and glutamate excitotoxicity [43], all of which combinedly contribute to the death of neurons due to imbalance between free radical production and degradation [83]. In AD, mitochondrial dysfunction plays a

potential role in cell death in which disturbance of energy metabolism leads to glutamate excitotoxicity [96,97]. The disturbed energy metabolism also enhances oxidative stress via excessive  $Ca^{++}$  influx in the cell leading to free radical production through p38 MAPK activation [15,16,98,99].

Moreover, p38 activation has been further demonstrated to be involved in cell death mechanisms in neuronal models. Treatment of neuronal cultures with arsenite, lipopolysaccharide, glutamate, amyloid beta, IL-1, TNF- $\alpha$ , ceramide and sulfidryl oxidizing agent results in p38 MAPK activation [38,100,101]. In AD brain, neurons become particularly sensitive to attack by free radicals. Various free

radicals like reactive oxygen species (ROS) and reactive nitrogen species (RNS) such as peroxy nitrite lead to lipid peroxidation, protein oxidation and DNA damages [102,103,104].

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### Abbreviations

Ad	Alzheimer's disease
A $\beta$	Amyloid beta
Ache	Acetylcholinesterase
Als	Amyotrophic lateral sclerosis
Ampa	A-amino-3-hydroxy-5-methyl-4-isoxazole propionate
App	Amyloid precursor protein
Ap-1	Activator protein-1
Ask1	Apoptosis-signal regulating kinase 1
Atf	Activating transcription factor
Atp	Adenosine triphosphate
Bmk1	Big mapk1
Csaids	Cytokine suppressive anti-inflammatory drugs
Chat	Cholineacetyltransferase
Chop	Caat enhancer binding protein homologous protein
Cox	Cyclooxygenase
Creb	Camp response element binding protein
Csf	Cerebrospinal fluid
Cns	Central nervous system
Cpcsea	Committee for the purpose of control and supervision of experiments on animals
Ddit3	Dna damage age inducible transcript3e
Elf-4e	Initiation factor-4e
Elk1	Ets like gene 1
Erks	Extracellular signal-regulated kinases
Fad	Familial alzheimer's disease
Fgf	Fibroblast growth factor
Gaba	Gamma amino butyric acid
Gsk	Glycogen synthase kinase
Gsh	Glutathione
Hsp	Heat shock protein
Hmg-14	High morbidity groups
H2b	Histone 2b
Il-1	Interleukin-1
Ial	Initial acquisition latency
Iaec	Institutional animal ethics committee
Inos	Inducible nitric oxide synthase
Igf	Insulin like growth factor
Icam	Intracellular adhesion molecule
Icv	Intracerebroventricular
Jnk	C-jun n-terminal kinase
Lox	Lipoxygenase
Lps	Lipopolysaccharide
Lts	leukotrienes
Ltp	long term potentiation
Ltd	long term depression
Madd	Mitogen activated kinase activating death domain protein
Mapk	Mitogen-activated protein kinase

Mapkk	Mitogen-activated protein kinase kinase
Mda	Malondialdehyde
Mhc-ii	Major histocompatibility complex-ii
Mk2	Map kinase-activated protein kinase 2 or mk2
Mef-2c	Myocyte enhanced factor 2c
Mekk	Mapk-extracellular signal-regulated kinase or mkkk
Mlk	Mixed lineage kinases
Mkk	Map kinase kinase
Mkkk	Map kinase kinase kinase or mekk
Mkp	Map kinase phosphatase
Mnk	Mapk signal integrating protein kinase
Mrna	Messenger ribonucleic acid
Msk	Mitogen and stress activated
Nfat	Nuclear factor of activated t-cells
Nf- $\kappa$ b	Nuclear factor kappa b
Nhe	Na <sup>+</sup> - h <sup>+</sup> exchanger isoform-1
Nft	Neuro fibrillary tangle
Nmda	N-methyl-d-aspartate
Phf	Paired helical filament
Prak	P38 regulating activated kinase
Pd	Parkinsonian diseases
Pdgf	Platelet derived growth factor
Pla2	Phospholipase a2
Pgs	Prostaglandins
Ps1	Presenelin1
Ros	Reactive oxygen species
Rns	Reactive nitrogen species
Rtl	Retention transfer latency
Sad	Sporadic alzheimer's disease
Sap1	Srf accessory protein
Sapk	Stress activated protein kinase
Stat	Signal transducer and activation of transcription 1a
Stz	Streptozotocin
Tak-1	Transforming growth factor-beta activated protein kinase 1
Tab1	Tak1-binding protein
Tao	Thousand and one kinase
Thr	Threonine
Tgf	Transforming growth factor
Tlr	Toll like receptor
Tyr	Tyrosine
Tnf	Tumor necrosis factor
Tnfr	Tumor necrosis factor receptor
Traf	Tumor necrosis factor receptor-associated factor
Vegf	Vascular endothelial growth factor

## References

1. Parihar MS, Hemani T. Alzheimer's diseases pathogenesis and therapeutic interventions. *J Clin Neur* 2004;11:456-467.
2. Perry G, Taddeo MA, Nunomura A, Zhu X, Zenteno-Savin T, Drew KL, Shimohama S, Avila J, Castellani JR, Smith MA. Comparative biology and pathology of oxidative stress in Alzheimer's and other neurodegenerative diseases: beyond damage and response. *Comp Biochem and Phys Part C* 2002;133:507-513.
3. Markesbery WR. Oxidative stress hypothesis in Alzheimer's diseases. *Free Rad Biol & Med* 1997;23:134-147.
4. Moore AH, O'Banion MK. Neuroinflammation and anti-inflammatory therapy for Alzheimer's diseases. *Adv Drug Del Rev* 2002;54:1627-1656.
5. Parachikova A, Agadjanyan MG, Cribbs DH, Blurton sJones, Perreau V, Rogers J, Beach TG, Cotman CW. Inflammatory changes parallel the early stages of Alzheimer's disease. *Neurol Bio Aging* 2007; **28**: 1821-33.
6. Selkoe DJ. Alzheimer's disease: Genes, Proteins and Therapy. *Physiol Rev* 2001; **81**: 741-65.
7. Daniels, W.M.U., Hendricks, J., Salie, R., Taljard, J.J.F. Role of the MAP-kinase superfamily in  $\beta$ -amyloid toxicity. *Mol. Brain. Dis* 2001; 16, 175-185.
8. Savage MJ, Lin YG, Ciallella JR, Flood DG, Scott RW. Activation of c-JUN-N-terminal kinase and p38 in an Alzheimer's disease model is associated with amyloid deposition. *J Neurosci* 2002;22:3376-3385.
9. Zhu X, Lee HG, Raina AK, Perry G, Smith MA. The role of mitogen activated protein kinase pathways in Alzheimer's diseases. *Neuro-Signals* 2002;11:270-281.
10. Alzheimer's diseases, Tauopathies and APP transgenic mice. *Neurotoxicity Res* 2004;6:1-8.
11. Jin Y, Yan EZ, Fan Y, Zong ZH, Qi ZM, Li Z. Sodium ferulate prevents amyloid-beta-induced neurotoxicity through suppression of p38 MAPK and upregulation of ERK-1/2 and AKT/protein kinase B in rat hippocampus. *Acta Pharmacol Sinica* 2005;26:943-951.
12. Jin W, Fan Y, Yan EH, Liu Z, Zong ZH, Qi ZM. Effects of sodium ferulate on amyloid-beta-induced MKK3/MK6-p38 MAPK-Hsp27 signal pathway and apoptosis in rat hippocampus. *Acta Pharmacol Sinica* 2006;27:1309-1316.
13. Eldik V, Mrak ER, Griffin WST. Interleukin-1 promotion of p38 MAPK overexpression in experimental animals and in Alzheimer's diseases: potential significance for tau protein phosphorylation. *Neurochem Int* 2001;39:341-348.
14. Munoz, L., Ranaivo, H.R., Roy, S.M., Hu, W., Craft, J.M., McNamara, L.K., Chico, L.W., Van, Eldik, L.J., Watterson, D. A novel p38 alpha MAPK inhibitor suppresses brain proinflammatory cytokine up-regulation and attenuates synaptic dysfunction and behavioral deficits in an Alzheimer's disease mouse model. *J. Neuroinflamm* 2007; 4, 1-14.
15. Molz S, Decker H, Dal-Cim T, Cremonez C, Cordova Fm, Leal RB, Tasca CI. Glutamate-induced toxicity in hippocampal slices involves apoptotic features and p38 MAPK signaling. *Neurochem Res* 2006;33:27-36.
16. Koistinaho, M., Koistinaho, J. Role of p38 and p44/42 mitogen activated protein kinases in microglia. *Glia* 2002;40, 175-183.
17. Griffin WS, Liu L, Li Y, Mrak RE, Barger SW. Interleukin-1 mediates Alzheimer and Lewy body pathologies. *J Neuroinflammation* 2006; 3: 1-9.
18. Bhat NR, Zhang P, Lee JC, Hogan EL. Extracellular signal-regulated kinase and p38 subgroups of mitogen-activated protein kinases regulate inducible nitric oxide synthase and tumor necrosis factor- $\alpha$  gene expression in endotoxin-stimulated primary glial cultures. *J Neurosci* 1998;18:1633-1641.
19. Bendotti C, Tortarolo M, Borsello T. Targeting Stress Activated Protein Kinase, JNK and p38, As a New Therapeutic Approach for Neurodegenerative Diseases. *Central Nervous System Agents In Medicine Chemistry* 2006; 6: 1-9.
20. Giovannini MG, Scali C, Prosperi C, Bellucci A, Vannucchi MG, Susanna R, Giancarlo P, Casamonti F.  $\beta$ -Amyloid-induced inflammation and cholinergic hypofunction in rat brain in-vivo: involvement of the p38 MAPK pathway. *Neurobiol Dis* 2002; 11: 257-74.
21. Zhu X, Mei M, Lee HG, Wang Y, Han J, Perry G, Smith MA. p38 activates mediated amyloid- $\beta$  cytotoxicity. *Neurochem Res* 2005;30:791-796.
22. Reynolds CH, Nebreda AR, Gibb GM, Utton MA, Anderton BH. Reactivating kinase/p38 phosphorylate  $\tau$  protein in vitro. *J Neurochem* 1997;69:191-198.
23. Hensly, K., Floyd, R.A., Zheng, N.Y., Nael, R., Robinson, K.A., Nguyen, Z., Pye, Q.N., Stewart, C.A., Geddes, J., Markesbery, W.R., Patel, E., Johnson, G.V.W., Bing, G. p38 MAPK is activated in the Alzheimer's disease brain. *J. Neurochem* 1999;72, 2053-2058.
24. Sun, A., Liu, M., Nguyen, X.V., Bing, G. p38 MAPK kinase is activated at early stages in Alzheimer's disease brain. *Exp. Neurol* 2003;183, 394-405.
25. Cuenda, A., Rousseau, S. p38 MAPK pathway regulation, function and role in human diseases. *Biochem. Biophys. Acta* 2007;1773, 1358-1375.
26. Evans, J.L., Goldfine, I.D., Maddux, B.A., Grodsky, G.M. Are oxidative stress-activated signaling pathways mediators of insulin resistance and  $\beta$ -cell dysfunction? *Diabetes* 2003; 52, 1-8.
27. Evans JI, Goldfine ID, Maddux BA, Grodsky GM. Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2 diabetes. *Endocrine reviews* 2008; **23**: 599-22.
28. Jiong C. The regulation and role of Stress-Activated protein Kinases ( p38 and JNK ) in neuronal death, Kuopio University Publications G. A-I. Virtanen Institute For Molecular Sciences 34. 2005. 79.
29. Lee, S.H., Park, J., Che, Y., Han, P.L., Lee, J.K. Constitutive activity and differential localization of p38 $\alpha$  and p38 $\beta$  MAPKs in adult mouse brain. *J. Neurosci* 2000; 60, 623-631.
30. Irving, E.A., Bamford, M. Role Of Mitogen And Stress Activated Kinases In Ischemic Injury. *Journal Of Cerebral Blood Flow & Metabolism* 2002;22: 631-647.
31. Rouse, J., Cohen, P., Trigon, S. A novel kinase cascade triggered by stress and heat shock that stimulates MAPKAP kinase-2 and phosphorylation of the small heat shock proteins. *Cell* 1994;78, 1027-1037.
32. Kyriakis, M.J., Joseph, A. Mammalian Mitogen Activated Protein Kinase Signal Transduction

- Pathways Activated By Stress And Inflammation. *Physiol. Rev* 2001; 81,807-69.
33. Han, J., Lee, J.D., Bibbs, L., Ulevitch, R.J. A MAP kinase targeted by endotoxin and hyperosmolarity in mammalian cells. *Sci* 1994; 265, 808-811.
  34. Lee, C.L., Laydon, J.T., McDonnell, P.C., Gallagher, T.F., Kumar, S., Green, D., McNulty, D., Blumenthal, M.J., Heys, J.R., Landvate, S.W. A protein kinase involved in the regulation of inflammatory cytokine biosynthesis. *Nature* 1994;72, 739-746.
  35. Jiang, Y.C., Chen, Z.L., Guo, W., Genger, J., Lin, S., Hann, J. Characterization of the structure and function of a new mitogen activated protein kinase (p38 $\beta$ ). *J. Biol. Chem* 1996;271, 17920-17926.
  36. Kumar, S., McDonnell, P.C., Gum, R.J., Hand, A.T., Lee, J.C., Young, P.R. Novel homologues of CSBP/p38 MAP kinase: activation, substrate specificity and sensitivity to inhibition by pyridinyl imidazoles. *Biochem. Biophys. Res. Commun* 1997; 23, 533-538
  37. Lechner, C., Zahyalka, M., Giot, M., Moller, N., Ullrich, A. ERK6, a mitogen activated protein kinase involved in C2C12 myoblast differentiation. *Proc. Natl. Acad. Sci. USA* 1996; 93, 4355-4359.
  38. Li, Z., Jiang, Y., Ulevitch, R.J., Han, J. The primary structure of p38 $\gamma$ : a new member of the p38 group of MAP kinases. *Biochem. Biophys. Res. Commun* 1996; 228, 334.
  39. Jiang, Y., Gram, H., Zhao, M., Feng, L., Di Padova, F., Ulevitch, R., Han, J. Characterization of the structure and function of the fourth member of p38 mitogen activated protein kinase. *J. Biol. Chem* 1997; 272, 30122-30128.
  40. Hanks, S.K., Hunter, T. The eukaryotic protein
  41. kinase superfamily: kinase (catalytic) domain structure and classification. *FASEB* 1999; J. 9, 576 596.
  42. Kumar, S., Boehm, J., Lee, J.C. p38 map kinases: key signaling Molecules as therapeutic targets for inflammatory diseases. *Nat. Rev* 2003; 2, 717-726.
  43. Bolshakov, V.Y., Carboni, L., Cobb, M.H., Siegelbaum, S.A., Belardetti, F. Dual MAP kinase pathways mediate opposing forms of long-term plasticity at CA3CA1. *Nature. Neurosci* 2000; 3, 1107 1112.
  44. Meikle K, Brecht S, Dorst A, herdegen T. Activity and expression of JNK1, p38 and ERK kinase, c-Jun N-terminal phosphorylation, and c-jun promoter binding in the adult rat brain following kainite induced seizures. *Neurosci* 1999; 91: 471-83.
  45. Pei JJ, Braak E, Braak H, Grundke-Iqbal I, Iqbal K, Winblad B, Cowburn RF. Localization of active forms of C-jun kinase (JNK) and p38 kinase in Alzheimer's disease brains at different stages of neurofibrillary degeneration. *J Alzheimers Dis* 2001; 3: 41-8.
  46. Alonso, G., Ambrosino, C., Jones, M., Nebreda, A.R. Differential activation of p38 mitogen-activated protein kinase isoforms depending on signal strength. *J. Biol. Chem* 2000; 275, 40641-40648.
  47. Guan, Z., Kim, J.H., Lomvardas, S., Holik, K., Kandel, E.R., Schwartz. p38 MAPK mediated both short-term and long term synaptic depression in Aplysia. *J. Neurosci* 2003; 23, 7317-7325.
  48. Brust, T.B., Cayabyab, F.S., Zhou, N., MacVicar, B.A. p38 mitogen-activated protein kinase contributes to adenosine A1 receptor-mediated synaptic depression in area CA1 of the rat hippocampus. *J. Neurosci* 2006; 26, 12427-12438.
  49. Goedert, M., Spillantini, M.G., Potier, M.C., Ulrich, J., Crowther, R.A. Cloning and sequencing of the cDNA encoding an isoform of microtubule associated protein tau containing four tandem repeats: differential expression of tau protein mRNAs in human brain. *EMBO* 1989; 8, 393-399.
  50. Goedert, M., Hasegawa, M., Jakes, R., Lawler, S., Cuenda, A., Cohen, P. Phosphorylation of microtubule-associated protein tau by stress-activated protein kinases. *FEBS. Lett* 1997; 409, 57-62.
  51. Reynolds, C.H., Betts, J.C., Blackstock, W.P., Nebreda, A.R., Anderton, B.H., Phosphorylation sites on tau identified by nano-electrospray mass spectrometry: differences in vitro between the mitogen-activated protein kinases ERK2, c-Jun N-terminal kinase and P38, and glycogen synthase kinase-3 beta. *J. Neurochem* 2000;74,1587-1595.
  52. Anderton, B.H., Betts, J., Blackstock, W.P., Brion, J.P., Chapman, S., Connell, J. Sites of phosphorylation in tau and factors affecting their regulation. *Biochem. Soc. Symp* 2001; 67, 73-80.
  53. Buee-Scherrer, V., Goedert, M. Phosphorylation of microtubule-associated protein tau by stress-activated protein kinases in intact cells. *FEBS. Lett* 2002; 515,151-154.
  54. Sheng, J.G., Ito, K., Skinner, R.D., Mrak, R.E., Rovnaghi, C.R., Van-Eldik, L.J. In vivo and in vitro evidence supporting a role for the inflammatory cytokine interleukin-1 as a driving force in Alzheimer pathogenesis. *Neurobiol. Aging* 1996; 117, 761-766.
  55. Ishihara, T., Hong, M., Zhang, B., Nakagawa, Y., Lee, M.K., Trojanowski, J.Q., Lee, V.M., Age dependent emergence and progression of a tauopathy in transgenic mice overexpressing the shortest human tau isoform. *Neuron* 1999; 24, 751-762.
  56. Ishihara, T., Zhang, B., Higuchi, M., Yoshiyama, Y., Trojanowski, J.Q., Lee, V.M. Age-dependent induction of congophilic neurofibrillary tau inclusions in tau transgenic mice. *Am. J. Pathol* 2001; 158, 555-562.
  57. Lewis, J., Dickson, D.W., Lin, W.L., Chisholm, L., Corral, A., Jones, G., 2001. Enhanced neurofibrillary degeneration in transgenic mice expressing mutant Tau and APP. *Sci* 2001; 293,1487 1491.
  58. Troy, C.M., Rabacchi, S.A., Xu, Z., Maroney, A.C., Connors, T.J., Shelanski, M.L., Greene, L.A. Beta-Amyloid-induced neuronal apoptosis requires c Jun N-terminal kinase activation. *J. Neurochem* 2001; 77, 157-164.
  59. Wei, W., Wang, X., Kusiak, J.W. Signaling events in amyloid beta-peptide-induced neuronal death and insulin like growth factor I protection. *J. Biol. Chem* 2002; 277,17649-17656.
  60. Savage, M., Gingrich, DE. Advances in the development of kinase inhibitor therapeutics for Alzheimer's disease. *Drug. Dev. Res* 2009; 70, 124-44.
  61. Zhu, X., Raina, A.K., Rottkamp, C.A., Aliev, G., Perry, G., Bux, H., Smith, M.A. Activation and redistribution of c-Jun N-terminal kinase/stress activated protein kinase in degenerating neurons in Alzheimer's disease. *J. Neurochem* 2001a;76, 435 441.
  62. Zhu, X., Rottkamp, C.A., Hartzler, A., Sun, Z., Takeda, A., Bux, H., Shimohama, S., Perry, G.,

- Smith, M.A. Activation of MKK6, an upstream activator of p38, in Alzheimer's disease. *J. Neurochem* 2001b;79, 311–318.
63. Morishima, Y., Gotoh, Y., Zieg, J., Barrett, T., Takano, H., Flavell, R., Davis, R.J., Shirasaki, Y., Greenberg, M.E. Beta-amyloid induces neuronal apoptosis via a mechanism that involves the c-Jun N-terminal kinase pathway and the induction of Fas ligand. *J. Neurosci* 2001; 21, 7551–7560.
64. Bozyczko-Coyne D, O'Kane TM, Wu ZL, Dobrzanski P, Murthy S, Vaught JL, Scott RW. CEP-1347/ KT-7515, an inhibitor of SAPK/JNK pathway activation, promotes survival and blocks multiple events associated with A $\beta$ -induced cortical neuron apoptosis. *J Neurochem* 2001; 77: 849-63.
65. Yan, S.D., Roher, A., Chaney, M., Zlokovic, B., Schmidt, A.M., Stern, D. Cellular cofactors potentiating induction of stress and cytotoxicity by amyloid beta-peptide. *Biochim. Biophys. Acta* 2000; 1502, 145–157.
66. Takeda, K., Ichijo, H. Neuronal p38 MAPK signalling: An emerging regulator of cell fate and function in the nervous system. *Genes. Cells* 2002; 7, 1099–1111.
67. Fillit, H., Ding, W.H., Buee, L., Kalman, J., Altstiel, L., Lawlor, B., Wolf-Klein, G. Elevated circulating tumor necrosis factor levels in Alzheimer's disease. *Neurosci. Lett* 1991; 129, 318–320.
68. Anderson, A.J., Cummings, B.J., Cotman, C.W. Increased immunoreactivity for Jun- and Fos-related proteins in Alzheimer's disease: Association with pathology. *Exp. Neurol* 1994; 125, 286–295.
69. Mielke, K., Herdegen, T. JNK and p38 stresskinases-degenerative effectors of signal transduction-cascades in the nervous system. *Prog. Neurobiol* 2000; 61, 45–60.
70. Belevsky, K., Maroun, M., Rosenblum, K. MAPK activation in the hippocampus in vivo is correlated with experimental setting. *Neurobiol. Learn. Mem* 2007; 88, 58–64.
71. Liang, Y.C., Huang, C.C., Hsu, K.S. A role of p38 mitogen-activated protein kinase in adenosine A1 receptor synaptic depotentiation in area CA1 of the rat hippocampus. *Mol. Brain* 2008; 1–13.
72. Armstrong, J.N., Brust, T.B., Lewis, R.G., MacVicar, B.A. Activation of presynaptic p2X<sub>7</sub>-like receptors depresses mossy fiber-CA3 synaptic transmission through p38 mitogen-activated protein kinase. *J. Neurosci* 2002; 22, 59438–59445.
73. Schulte, G., Fredholm, B.B. Signaling from adenosine receptors to mitogen-activated protein kinase. *Cell. Signal* 2003; 15, 813–827.
74. Guan, Z., Buckman, S.Y., Pentland, A.P., Templeton, D.J., Morrison AR. Induction of cyclooxygenase-2 by the activated MEKK1/SEK1/ MKK4/p38 mitogen-activated protein kinase pathway. *J. Biol. Chem* 1998; 273, 12901–12908.
75. Coogan, A., Lynch, M.A., O'Connor, J.J. The p38 MAP kinase inhibitor, SB203580, antagonizes the effect of IL-1 $\beta$  on LTP in rat dentate gyrus. *Neurosci* 1997; 93, 57–59.
76. O'Donnell, E., Vereker, E., Lynch, M.A. Age-related impairment in LTP is accompanied by enhanced activity of stress-activated protein kinases: analysis of underlying mechanisms. *Eur. J. Neurosci* 2000; 12, 345–352.
77. Vereker, E., O'Donnell, Lynch, M.A. The inhibitory effect of interleukin-1 $\beta$  on long-term potentiation is coupled with increased activity of stress-activated protein kinases. *J. Neurosci* 2000; 20, 6811–6819.
78. Liang, K.C., McGaugh, J.L., Martinez, J.R., Jensen, R.A., Vasquez, B.J., Messing, R.B. Post-training amygdaloid lesions impair retention of an inhibitory avoidance response. *Behav. Brain. Res* 1982; 4, 237–249.
79. Biessels, G.J., Kamal, A., Ramakers, G.M., Urban, I.J.A., Spruijt, B.M., Erkelens, D.W., Gispen, W.H. Place learning and hippocampal synaptic plasticity in streptozotocin-induced diabetic rats. *Diabetes* 1996; 45, 1259–1266.
80. Chabot, C., Massicotte, G., Milot, M., Trudeau, F., Gange, J. Impaired modulation of AMPA receptors by calcium-dependent processes in streptozotocin-induced diabetic rats. *Brain. Res* 1997; 768, 249–256.
81. Kamal A, Biessels GJ, Urban IJA, Gispen WH. Hippocampal synaptic plasticity in streptozotocin-diabetic rats: impairment of long-term potentiation and facilitation of long term depression. *Neurosci* 1999; 90: 737-45.
82. Kamal A, Ramakers GMJ, Biessels GJ, Gispen WH. Effects of phorbol ester and cyclosporine A on hippocampal synaptic plasticity in streptozotocin-induced-diabetic rats: reduced sensitivity to phorbol esters. *Neurosci Lett* 2003; 339: 45-8.
83. Culbert AA, Skaper SD, Howlett DR, Evans NA, Facci L, Soden PE, Seymour Zm, Florence Guillot, Gaestel M, Richardson JC. MAPK-activated protein kinase 2 deficiency in microglia inhibits proinflammatory mediator release and resultant neurotoxicity. *J Biol Chem* 2006; 281:23658-67.
84. Chaparro-Heurta, V., Flores-Soto, M.E., Gudino-Caberera, G., Rivera-Cervantes, M.C., Bitzer-Quintero, O.K., Beas-Zarate, C. Role of p38 MAPK and pro-inflammatory cytokines expression in glutamate-induced neuronal death of neonatal rats. *Int. J. Neurosci* 2008; 26, 487–495.
85. Walton, K.M., DiRocco, R., Bartlett, B.A., Koury, E., Marcy, V.R., Jarvis, B., Schaefer, E.M., Bhat, R.V. Activation of p38MAPK in microglia after ischemia. *J. Neurochem* 1998; 70, 1764–1767.
86. Quintanilla, R., Orellana, D.I., Gonzalez-Billault, C., Maccioni, R.B. Interleukin-6 induces Alzheimer-type phosphorylation of tau protein by deregulating the cdk5/p35 pathway. *Exp. Cell. Res* 2004; 295, 245–257.
87. Yoo, B.K., Choi, J.W., Shin, S.L., Jeon, S.J., Park, S.J., Cheong, J.H., Han, S.Y., Ryu, J.R., Song, M.R., Ko, K.H. Activation of p38 MAPK induced peroxynitrite generation in LPS plus IFN- $\gamma$  stimulated rat primary astrocytes via activation of iNOS and NADPH oxidase. *Neurochem. Int* 2008; 52, 1188–1197.
88. Beyaert, R., Cuenda, A., Vanden, B.W., Plaisance, S., Lee, J.C., Haegeman, G., Cohen, P., Fires, W. The p38/ERK mitogen-activated protein kinase pathway regulates interleukin-6 synthesis response to tumor necrosis factor. *EMBO* 1996; 15, 1914–1923.
89. Hale, K.K., Trollinger, D., Rihanek, M., Manthey, C.L. Differential expression and activation of p38 mitogen-activated protein kinase  $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$  in inflammatory cell lineages. *J. Immunol* 1999; 162, 4246–4252.



90. Suzuki, T., Hide, I., Ido, K., Kohsaka, S., Inoune, K., Nakata, Y. Production and release of neuroprotective tumor necrosis factor by P2X7 receptor activated microglia. *J. Neurosci* 2004; 24, 1-7
91. Butterfield, D., Perluigi, M., Sultana, R. Oxidative stress in Alzheimer's disease brain: New insights from redox proteomics. *Eur. J. Pharmacol* 2006; 545, 39-50.
92. Hsieh, C.C., Papaconstantinu, J. The effect of aging on p38 signaling pathway activity in the mouse liver and in response to ROS generated by 3-nitropropionic acid. *Mech. Aging. Dev* 2002; 123, 1423-1435.
93. Nolan, Y., Vereker, E., Lynch, A.M., Lynch, M.A. Evidence that lipopolysaccharide-induced cell death is mediated by accumulation of reactive oxygen species and activation of p38 in rat cortex and hippocampus. *Exp. Neurol* 2003; 184, 794-704.
94. Choe, E.S., McGinty, J.E. N-methyl-D-Aspartate receptors and p38 mitogen activated protein kinase are required for cAMP-dependent cyclase response element binding protein and ELK-1 phosphorylation in the striatum. *Neurosci* 2000; 101, 601-617.
95. Carracedo, A., Egia, A., Guzman, M., Velasco, G., 2006. p38 upregulation sensitizes astrocytes to oxidative stress. *FEBS. Lett* 2006; 580, 1571-1575.
96. Dewil, M., Cruz, V.F.D., Bosch, L.V.D., Robberecht, W. Inhibition of p38 mitogen activated protein kinase activation and mutant SOD1<sup>G93A</sup> - induced motor neuron death. *Neurobiol. Dis* 2007; 26, 332-341.
97. Barone, F.C., Irving, E.A., Ray, A.M., Lee, J.C., Kasis, S., Kumar, S., Badger, M.A., White, R.F., Mcvey, M.J., Legos, J.J., Erhardt, J.A., Nelson, H.A., Ohlstein, E.H., Hunter, A.J., Ward, K., Smith, B.R., Adams, J.L., Parsons, A.A. SB239063, a second-generation p38 mitogen-activated protein kinase inhibitor, reduces brain injury and neurological deficits in cerebral focal ischemia. *J. Pharmacol. Exp. Therap* 2001; 296, 312-321.
98. Piao, C.S., Kim, J.B., Han, P.L., Lee, J.K. Administration of the p38 MAPK inhibitor SB203580 affords brain protection with a wide therapeutic window against focal ischemic insult. *J. Neurosci. Res* 2003; 73, 537-544.
99. Yang, L., Mao, L., Tang, Q., Samdani, S., Liu, Z., Wang, J.Q. A novel Ca<sup>+</sup>-independent signaling pathway to extracellular signal-regulated protein kinase by coactivation of NMDA receptors and metabotropic glutamate receptor 5 in neurons. *J. Neurosci* 2004; 24, 10846-10857.
100. Guo, G., Bhat, N.R. p38 $\alpha$  MAP kinase mediated hypoxia-induced motor neuron cell death: A potential target of minocycline's neuroprotective action. *Neurochem. Res* 2007; 32, 216-266.
101. Honda, S., Nakajima, K., Nakamura, Y., Imai, Y., Kohsaka, S. Rat primary cultured microglia express glial cell line-derived neurotrophic factor receptors. *Neurosci. Lett* 1999; 275, 203-206.
102. Liva, S.M., Kahn, M.A., Dopp, J.M., De-Vellis, J. Signal transduction pathways induced by GM-CSF in microglia: significance in the control of proliferation. *Glia* 1999; 26, 344-352.
103. Akiyama, H., Barger, S., Barnum, S., Bradt, B., Bauer, J., Cole, G.M., Cooper, N.R., Eikelenboom, P., Emmerling, M., Fiebich, B.L., Finch, C.E., Frautschy, S., Griffin, W.S., Hampel, H., Hull, M., Landreth, G., Lue, L., Mrak, R., Mackenzie, I.R., McGeer, P.L., O'Banion, M.K., Pachter, J., Pasinetti, G., Plata Salaman, C., Rogers, J., Rydel, R., Shen, Y., Streit, W., Strohmeyer, R., Tooyoma, I., Van-Muiswinkel, F.L., Veerhuis, .R., Walker, D., Webster, S., Wegrzyniak, B., Wenk, G., Wyss-Coray, T. Inflammation and Alzheimer's disease. *Neurobiol. Aging* 2000; 21, 383-321.
104. Singh, R.P., Sharad, S., Kapur, S. Free radicals and oxidative stress in neurodegenerative diseases: Relevance of dietary antioxidants. *J. Ind. Acad. Clin. Med* 2004; 5, 218-225.